

# Selinexor, a Selective Inhibitor of Nuclear Export (SINE), shows enhanced activity in combination with PD-1/PD-L1 blockade in syngeneic murine models of colon cancer and melanoma

The James

THE OHIO STATE UNIVERSITY  
COMPREHENSIVE CANCER CENTER

Matthew R. Farren<sup>1</sup>, Reena Shakya<sup>1</sup>, Rebecca Hennessey<sup>1</sup>, Thomas Mace<sup>1</sup>, Jennifer Yang<sup>1</sup>, Omar Elnaggar<sup>1</sup>, Gregory Young<sup>1</sup>, Yosef Landesman<sup>2</sup>, Robert Carlson<sup>2</sup>, Sivan Elloul<sup>2</sup>, Marsha Crochiere<sup>2</sup>, Christin Burd<sup>1</sup>, Gregory B. Lesinski<sup>1</sup>

<sup>1</sup>The Ohio State University, Columbus, OH  
<sup>2</sup>Karyopharm Therapeutics, Newton, MA

## Abstract

Exportin-1 (XPO1) is a nuclear export protein with >220 cargo proteins, including tumor suppressors and cell cycle modulators. Selinexor (KPT-330), an oral XPO1 inhibitor, has been administered to >1000 cancer patients in Phase I and II trials, with evidence of efficacy and tolerability. Selinexor blocks nuclear export of NFAT1c, STAT1 and STAT3, which have been implicated in regulation of the inhibitory T cell receptor PD-1 and its ligand, PD-L1. We hypothesized that selinexor would upregulate T cell checkpoint molecule expression, and that combination treatment with anti-PD-1 or anti-PD-L1 antibodies would thereby enhance the ability of selinexor to elicit anti-tumor activity. Selinexor increased PD-1 gene expression by ~2-fold in normal lymphocytes and induced PD-L1 gene expression in tumor cell lines. Mice bearing syngeneic colon (C26) tumors and treated with selinexor ± anti-PD-1 antibodies for 2 weeks demonstrated a significant reduction in tumor growth rate ( $p < 0.05$ ), while monotherapy with either agent had no significant effect on tumor growth. Similar results were obtained in mice bearing syngeneic B16F10 tumors, whereby combined treatment with selinexor + anti-PD-1 antibodies was superior to either single agent ( $p < 0.034$ ). Combined therapy of mice bearing B16F10 tumors with selinexor and anti-PD-1 antibodies was similarly effective, with significantly smaller tumors at the study endpoint ( $p < 0.001$ ). Importantly, no weight loss or signs of toxicity were evident in any *in vivo* study. Immunophenotypic analysis of splenocytes revealed that combined therapy with selinexor + anti-PD-1/anti-PD-L1 antibodies significantly increased the frequency of NK cells ( $p < 0.045$ ) and significantly increased the frequency of TH1 T cells ( $p < 0.016$ ) as compared to vehicle treated mice. Interestingly, combining selinexor with anti-PD-L1 antibodies significantly decreased the proportion of PD-L1<sup>+</sup> MDSC, dendritic cell, NK cell, and T cell ( $p = 0.029$ ,  $p = 0.004$ ,  $p = 0.019$ ,  $p = 0.005$ , respectively). Anti-PD-1 antibody treatment did not alter PD-1 or PD-L1 expression in the tumor or individual splenic cell populations. However, ~40% of splenic T cells expressed PD-1 in all treatment groups. These data indicate that the efficacy of selinexor may be enhanced by disrupting the pre-existing PD-1/PD-L1 signaling in effector cells (T and NK cells). Altogether, these data suggest that the efficacy of selinexor in combination with anti-PD-1 or anti-PD-L1 in mouse syngeneic tumor models may be due to both disrupting immunosuppressive PD-1/PD-L1 signaling and increasing the frequency of potentially tumor reactive NK cells and TH1 T cells. This provides a rational basis for this treatment combination as a novel therapeutic approach for advanced cancer.

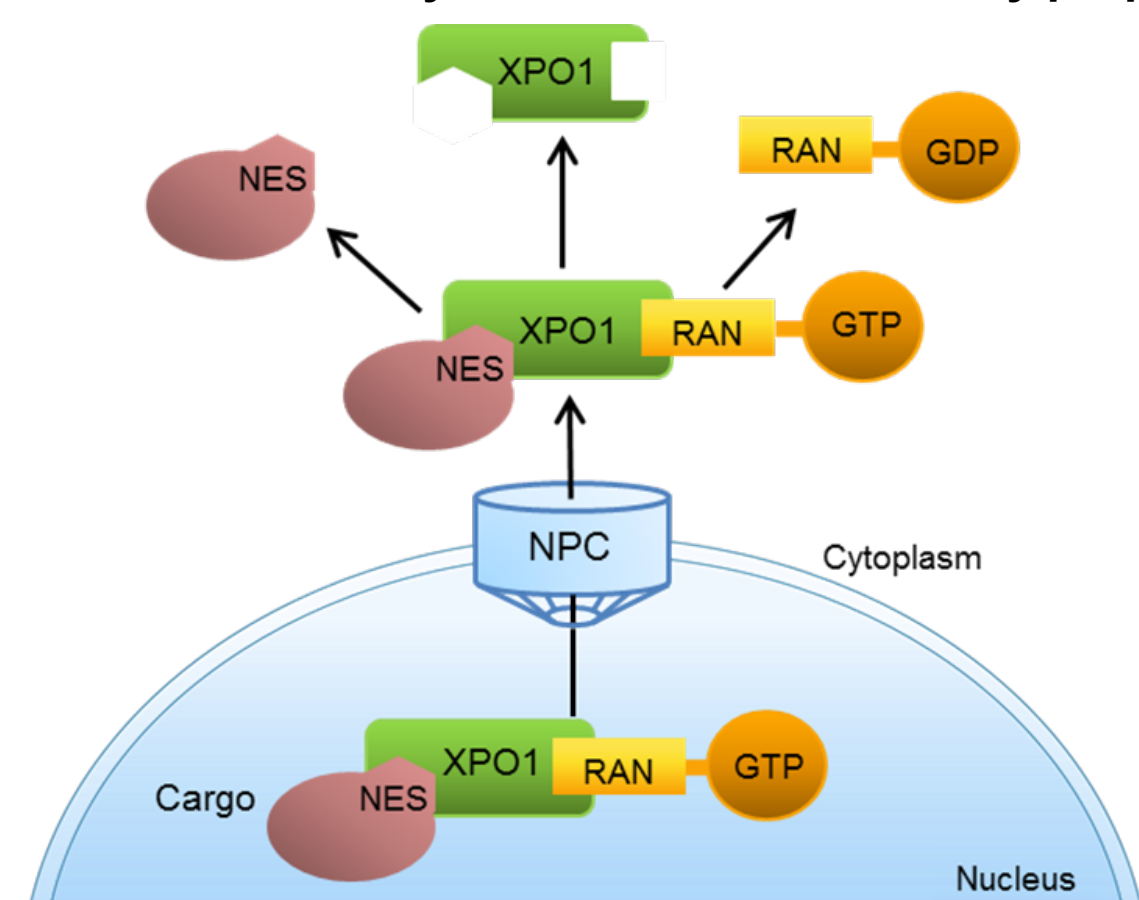
## Combining nuclear export inhibition with immunotherapy in melanoma

### Immunotherapy in melanoma

- Immune checkpoint blockade (ipilimumab, nivolumab) has clinical efficacy against metastatic melanoma, having elicited significant and sustained responses in a proportion of patients.
- While promising, majority of patients still progress in < 2 years following immunotherapy.

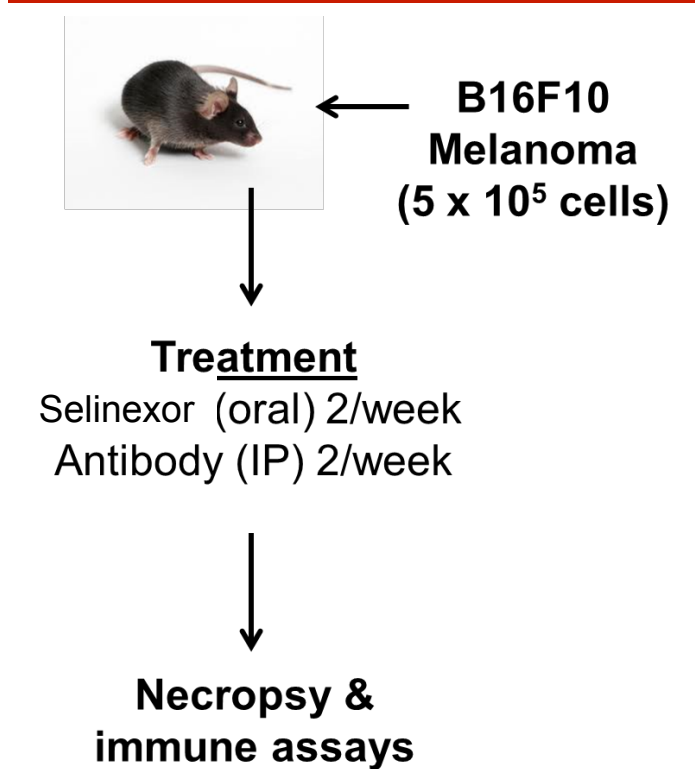
### Nuclear export inhibition in melanoma

- The nuclear export protein exportin-1 (XPO1) is upregulated during melanoma progression
- XPO1 transport >220 identified cargo proteins including tumor suppressors and cell cycle modulators
- Selinexor (KPT-330), a Selective Inhibitor of Nuclear Export (SINE) compound, inhibits XPO1
- In Yang *et al.* (*PLoS ONE*, 2014), we demonstrated this drug has direct anti-tumor activity in melanoma
- In addition to its antitumor activity, selinexor inhibits nuclear export of NFAT1c, STAT1, and STAT3 and may have immunomodulatory properties



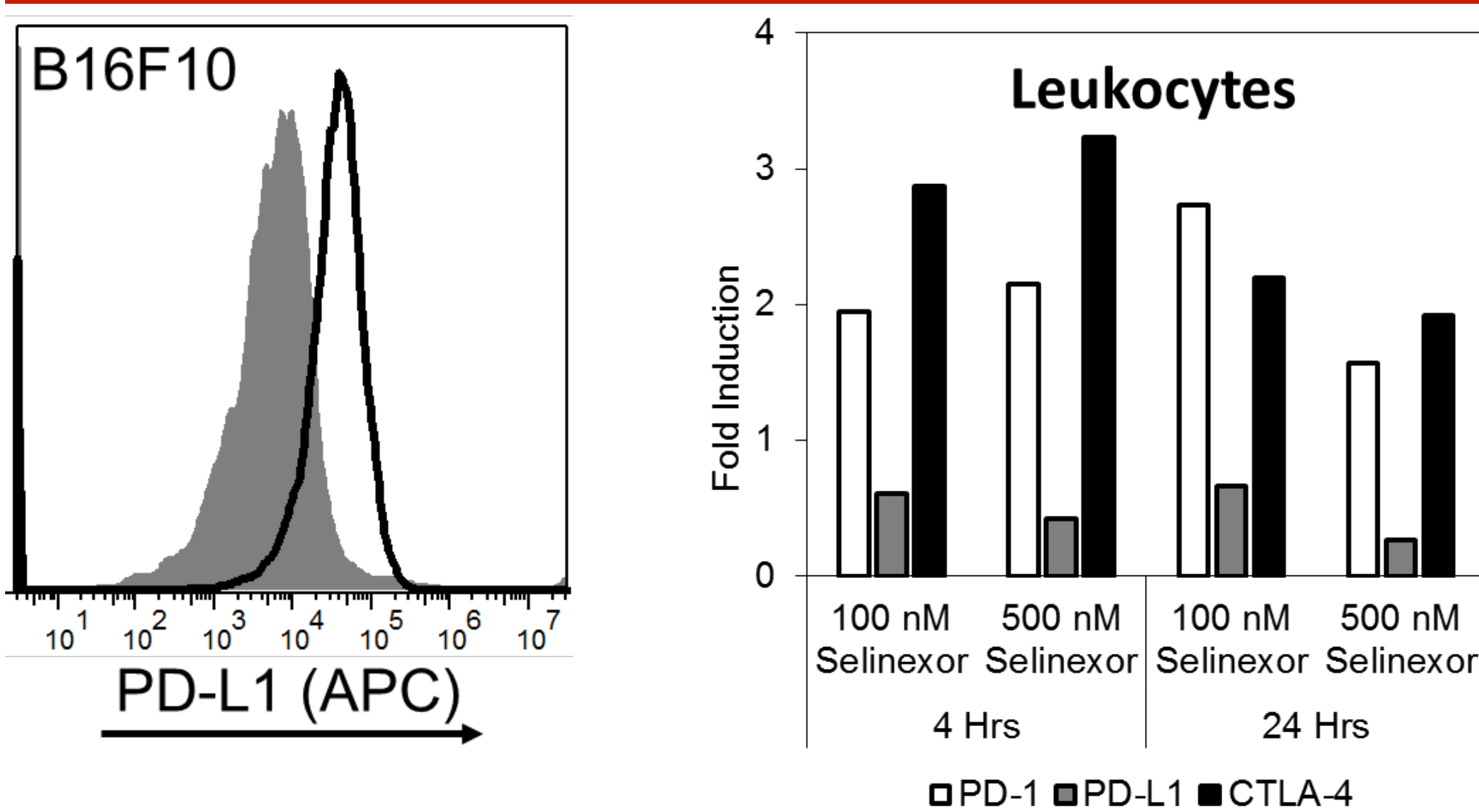
**Hypothesis:** Selinexor leads to direct anti-tumor activity but also upregulate T cell checkpoint molecule expression. Thus, combination treatment with anti-PD-1 or anti-PD-L1 will synergize with selinexor to control tumor growth

## Experimental design



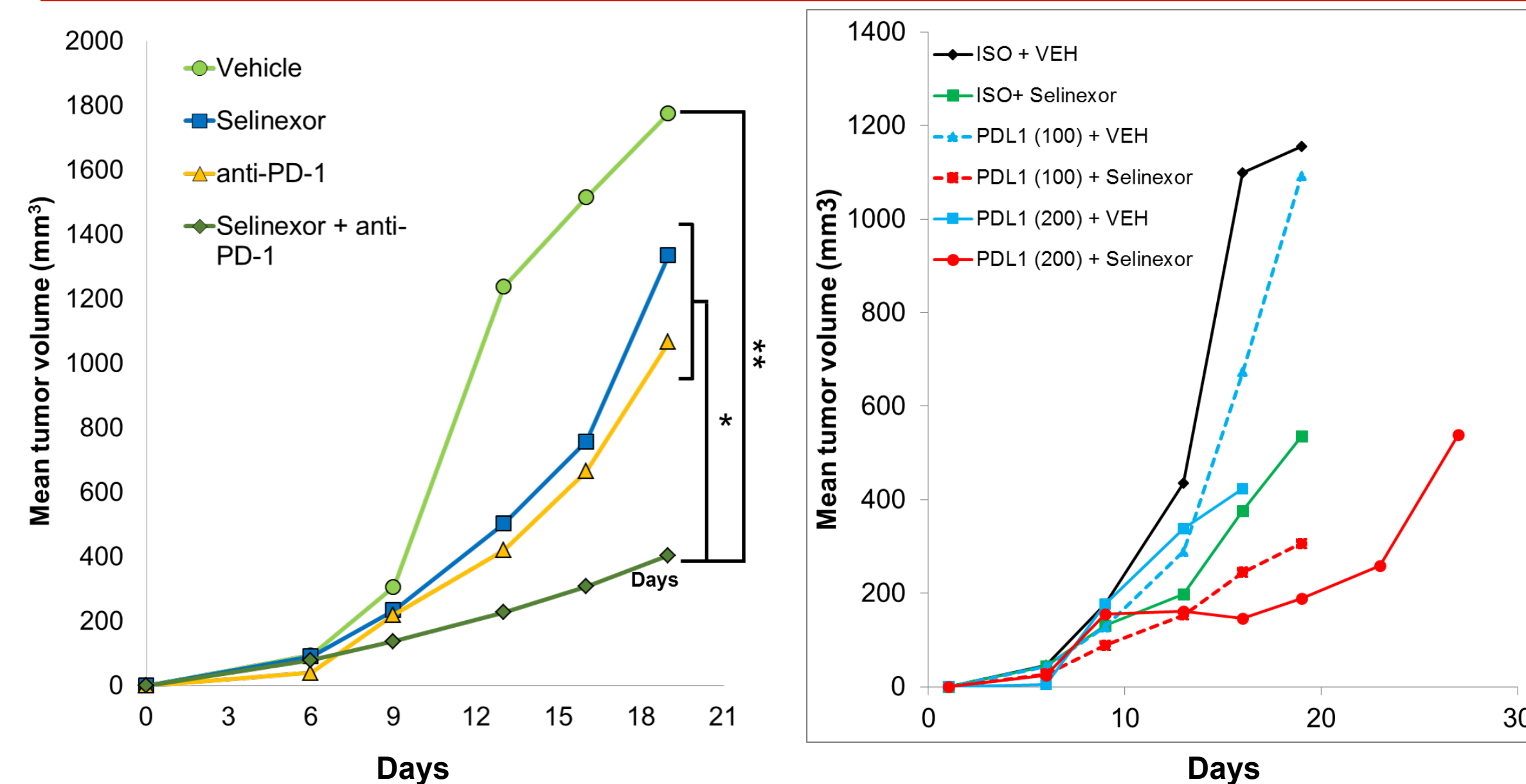
**Figure 1. Experimental Design:** 5.0 x 10<sup>5</sup> B16F10 melanoma cells were implanted subcutaneously into (immunocompetent) female C57BL/6 mice. When tumors became palpable, mice were treated with 15 mg/kg selinexor (orally) and 100-200 µg of the relevant antibody or isotype control (i.p.). All treatments were twice per week and continued until control tumors > 1500 mm<sup>3</sup>. Mice were then euthanized and tumor tissue, splenocytes, and plasma analyzed.

## Selinexor induces immune checkpoint molecule expression



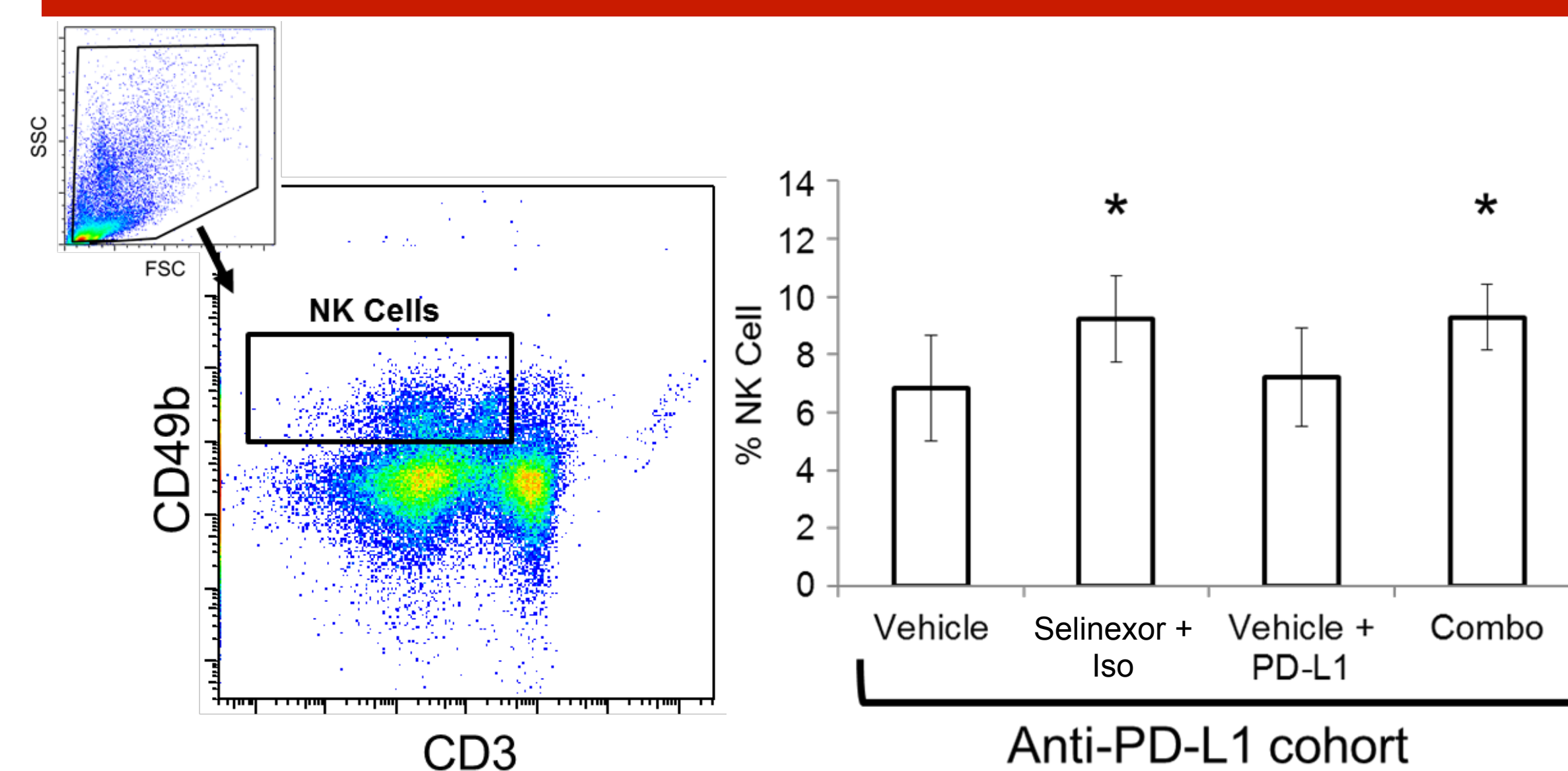
**Figure 2. Melanoma cells express PD-L1 at baseline and selinexor induces increased immune cell expression of PD-1, and CTLA-4** (left) PD-L1 expression on B16F10 was assessed by flow cytometry. B16F10 expressed PD-L1 at baseline (grey histogram: isotype control, black histogram: anti-PD-L1 stained). Selinexor incubation (up to 1 µM for 72 hours) did not further induce PD-L1 expression (data not shown). (right) Human leukocytes from healthy donors were incubated with 100-500 nM selinexor or diluent control for 4-24 hours and T cell checkpoint molecule expression assessed by qRT-PCR. Selinexor strongly induced PD-1 and CTLA-4 expression, while marginally suppressing PD-L1 expression in human leukocytes.

## PD-1/PD-L1 blockade + XPO1 inhibition by selinexor slows melanoma tumor growth



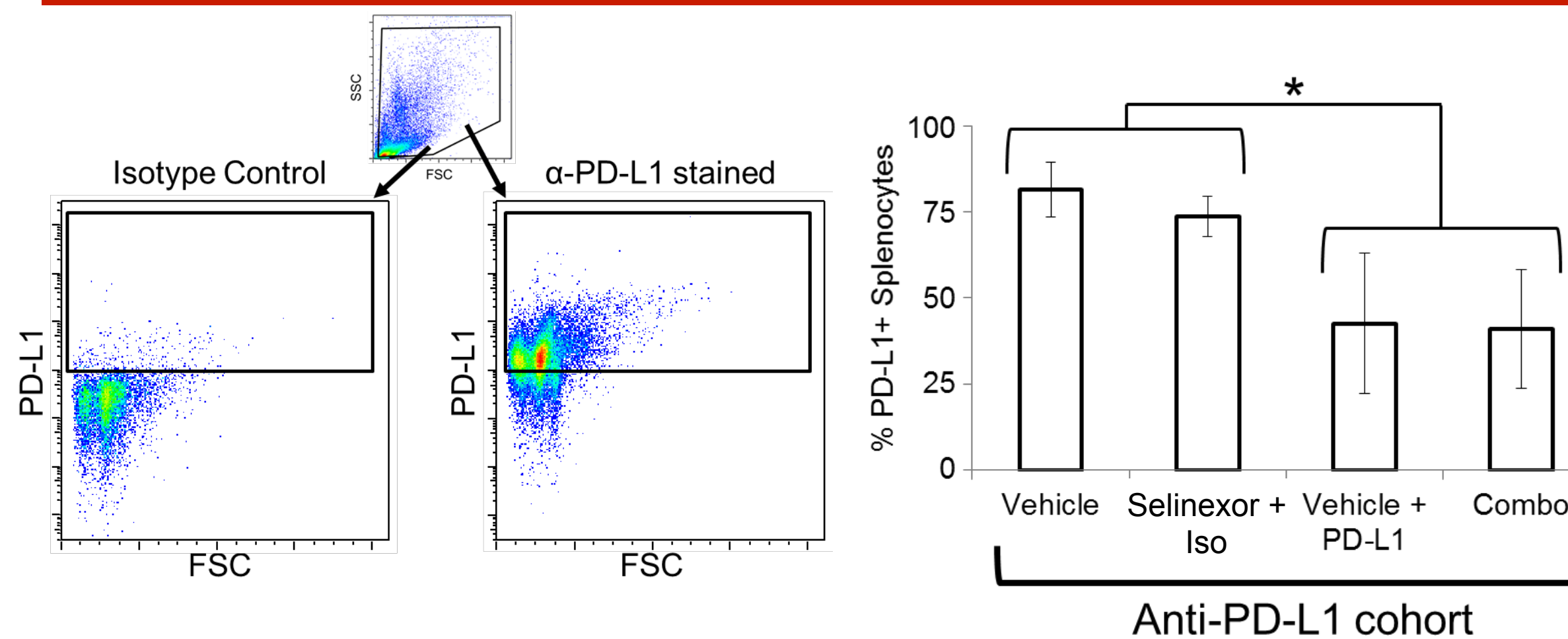
**Figure 3. Combination therapy with PD-1/PD-L1 blockade + selinexor exerts superior anti-tumor activity to either therapy alone.** Mice bearing B16F10 tumors were treated as described above with selinexor ± α-PD-1 or α-PD-L1 antibodies (PD-1/PD-L1 blockade), or diluent vehicle / isotype controls (VEH, ISO), twice weekly when tumors became palpable. Mice were euthanized when tumors grew to > 1500<sup>3</sup> or ulcerated. Lines are median tumor volume, n = 6 mice per group. \*  $p < 0.05$ , \*\*  $p < 0.01$ .

## Selinexor increases the frequency of NK cells in the spleen



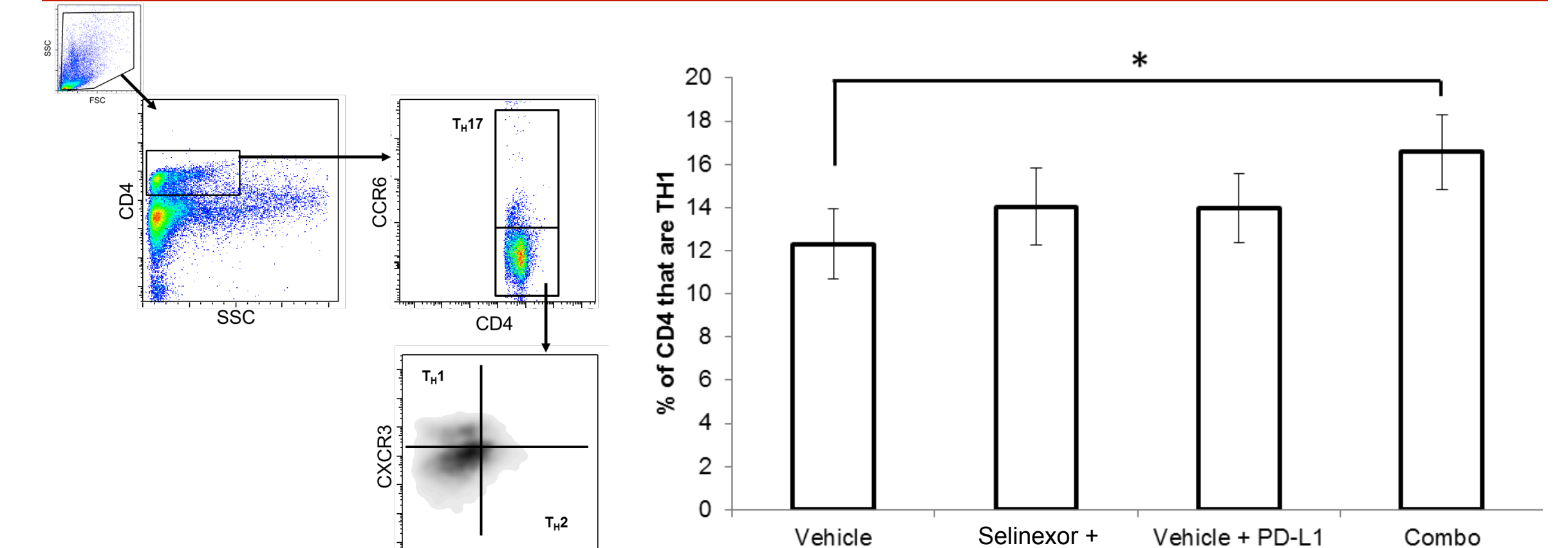
**Figure 5. Therapy with selinexor ± PD-1/PD-L1 blockade induces increased NK cell frequency.** (left) Gating strategy: NK cells were identified by gating CD3<sup>+</sup> CD49b<sup>+</sup> cells. (right) Selinexor induced an increased frequency of this potentially cytotoxic and anti-tumor cell type. n = 5-6 mice per group. \*  $p < 0.05$ .

## PD-L1 blockade decreases the frequency of PD-L1<sup>+</sup> splenocytes



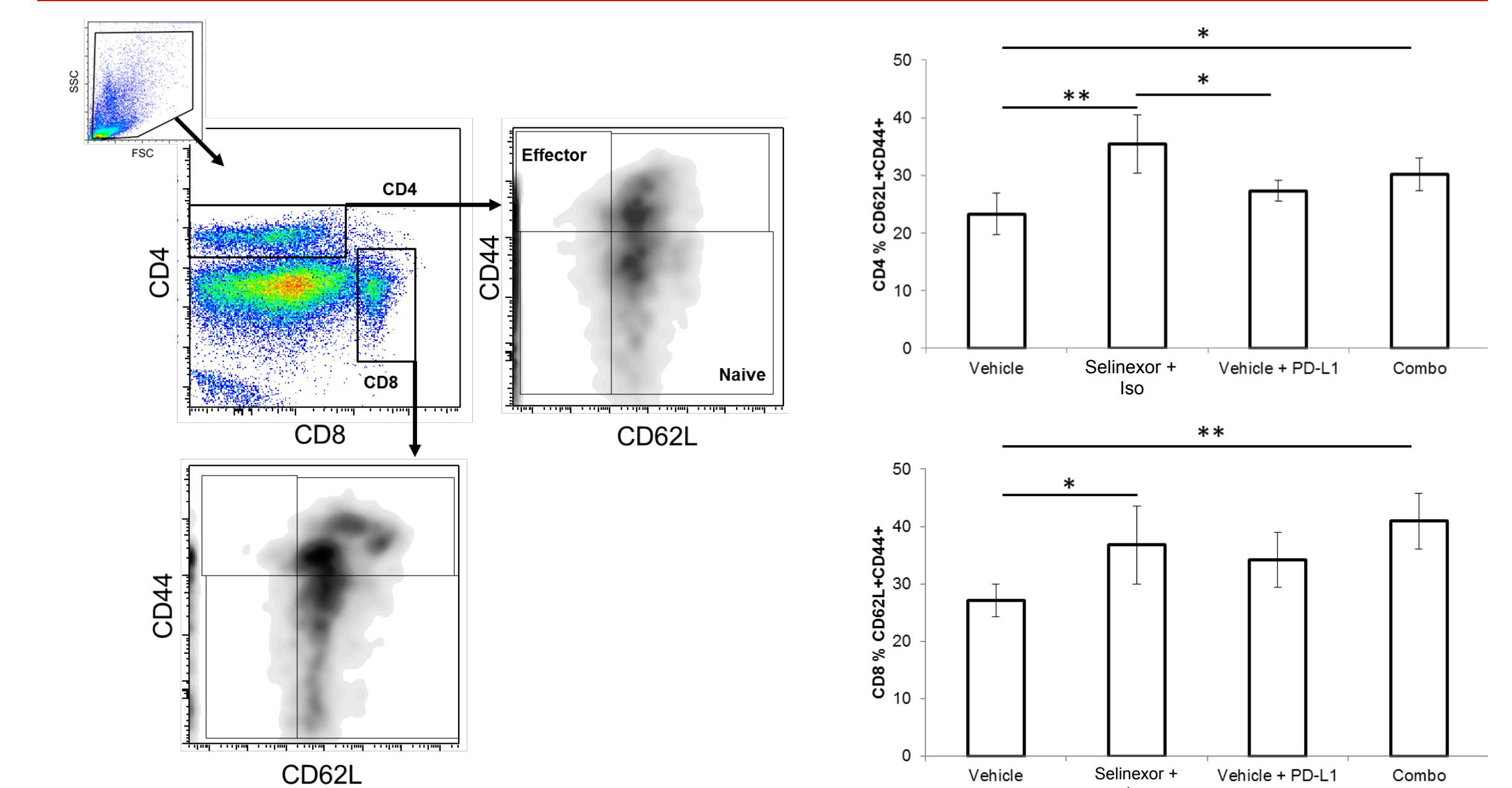
**Figure 6. Therapy with PD-L1 blockade ± selinexor decreased surfaced expression of PD-L1 on bulk splenocytes.** (left) Gating & validation strategy: PD-L1 expression on bulk splenocytes was assessed by flow cytometry. Expression was determined relative to isotype-control stained cells. To verify that decreased fluorescence was a result of decreased PD-L1 expression and not an artifact resulting from interference between the treatment α-PD-L1 antibody and the fluorescently labeled antibody, residual unlabeled (treatment) α-PD-L1 staining was assessed by staining "unlabeled" cells with a secondary antibody conjugated to FITC. (right) α-PD-L1 alone or in combination with selinexor significantly decreased PD-L1 expression on bulk splenocytes. n = 5-6 mice per group. \*  $p < 0.05$ .

## Selinexor + PD-L1 blockade induces splenic TH1 differentiation



**Figure 7. Combination therapy significantly increases TH1 differentiation.** (left) Gating strategy: Helper T cell phenotypes were determined based on CD4<sup>+</sup> T cell expression of CXCR3 (T<sub>H1</sub>), CCR4 (T<sub>H2</sub>), and CCR6 (T<sub>H17</sub>). (right) Combo therapy with selinexor + PD-L1 blockade significantly increased the frequency of cells with a T<sub>H1</sub> phenotype. n = 5-6 mice per group. \*  $p < 0.05$ .

## Selinexor increases *in vivo* splenic T cell activation



**Figure 8. Combination therapy significantly increases CD4<sup>+</sup> and CD8<sup>+</sup> T cell activation *in vivo*.** (left) Gating strategy: T cell activation status was assessed by first gating on CD4<sup>+</sup> or CD8<sup>+</sup> cells and then assessing CD44 and CD62L levels. CD62L<sup>+</sup> CD44<sup>-</sup> - Naive phenotype; CD62L<sup>+</sup> CD44<sup>+</sup> - early activation and central memory phenotype; CD62L<sup>-</sup> CD44<sup>+</sup> - Effector phenotype. Helper T cell phenotypes were determined based on CD4<sup>+</sup> T cell expression of CXCR3 (T<sub>H1</sub>), CCR4 (T<sub>H2</sub>), and CCR6 (T<sub>H17</sub>). Selinexor treatment significantly increased the proportion of T cells with early activated/central memory phenotype (right), as well as increasing effector phenotype and decreasing naive phenotype (data not shown). n = 5-6 mice per group. \*  $p < 0.05$ .

## Conclusions

- Combination of selinexor + PD-1 or PD-L1 blockade exerts considerable anti-tumor activity in an aggressive murine melanoma model.
- Combination of selinexor + PD-1 or PD-L1 blockade shows significant immunomodulatory activity, inducing changes in the frequency and phenotype of immune cell populations including:
  - Increased frequency of NK cells
  - Induced frequency of differentiated TH1 cells
  - Increased frequency of activated T-cells
- These results support the future clinical evaluation of the combination of selinexor + PD-1/PD-L1 blockade in melanoma patients

## Acknowledgements

We thank the OSU CCC Analytical Cytometry, Biostatistics, and Target Validation Shared Resources.

This project was supported in part by NIH Grants 5T32CA009338-34 (Caligiuri), 5T32CA090223-13 (Carson), 1R01 CA169363-01 (Lesinski), a fellowship from the Pelotonia Foundation, funding support from Karyopharm Therapeutics, and the generous support of The Siegle Fund for Melanoma Research and The Gill Fund for Melanoma Research.